

**Figure 2.** Schematic energy surface relating II and III. Height of singlet maximum (39 kcal/mol) is taken from J. P. Chesick, *J. Am. Chem. Soc.*, **84**, 3250 (1962). The cross hatched area is the barrier through which the system tunnels via a normal mode consisting mainly of a ring bending vibration.

ture range studied with a difference between the fast and slow component of approximately a factor of two. This behavior has precedent<sup>10</sup> and is presumably due to an ensemble of slightly different host sites, differing in their restraining power for the out-of-plane motion accompanying the formation of III. In addition both components of the rate are essentially temperature independent between 1.3 and 20°K! Only above 20°K does the disappearance of II begin to show a significant temperature dependence. The rate of II-*d*<sub>8</sub> at 5.5°K is too slow to study. This behavior is characteristic for quantum mechanical tunneling through a low barrier.<sup>11</sup> Estimates of the shape of the barrier are made difficult by the fact that the extremely small transmission coefficient ( $10^{-15}$  to  $10^{-16}$  sec<sup>-1</sup>) has a contribution from the spin forbiddenness of the reaction. Nevertheless, considering the large reduced mass of the tunneling groups, it is hard to see how the barrier height could exceed 2 kcal/mol.<sup>12</sup> A more detailed study of the kinetic behavior above 20°K on II and II-*d*<sub>8</sub> should yield a better estimate.

The results reported here are of interest in connection with the problem of whether the singlet surface has a minimum associated with structure II. While thermochemical calculations indicate such a well to be 13 kcal/mol deep<sup>13</sup> our results appear to be more consistent with a surface as indicated in Figure 2 where there is no minimum with singlet character. This conclusion is in line with quantum mechanical calculations on trimethylene.<sup>14</sup>

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## The Nonparticipation of $\alpha,\beta$ -Dehydrovalinyl Intermediates in the Formation of $\delta$ -(L- $\alpha$ -Aminoamidipyl)-L-cysteinyl-D-valine

Sir:

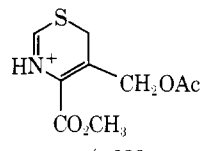
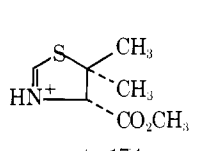
The tripeptide  $\delta$ -( $\alpha$ -aminoamidipyl)cysteinylvaline was first isolated from *Penicillium chrysogenum* in 1960,<sup>1</sup> but the absolute configurations of its constituent amino acids were not determined. Subsequently, a similar noncyclic peptide was isolated from *Cephalosporium acremonium*, characterized as  $\delta$ -(L- $\alpha$ -aminoamidipyl)-L-cysteinyl-D-valine (ACV)<sup>2</sup> and shown to be formed from  $\delta$ -(L- $\alpha$ -aminoamidipyl)-L-cysteine and L-valine, but not D-valine, in cell extracts.<sup>3,4</sup> More recently, ACV was shown to be a precursor of penicillin N as manifested by its incorporation into the latter  $\beta$ -lactam antibiotic by a cell-free system obtained from protoplasts of *C. acremonium*.<sup>5</sup>

Several recent publications<sup>6,7</sup> report the asymmetric incorporations of chirally labeled methylvaline-*methyl*-<sup>13</sup>C and methylvalines-*methyl*-*d*<sub>3</sub> into  $\beta$ -lactam antibiotics. We present here the results of biosynthetic studies with (2*S*,3*S*)-methylvaline-<sup>15</sup>N,3-*methyl*-*d*<sub>3</sub> and experimental evidence for the nonparticipation of  $\alpha,\beta$ -dehydrovalinyl intermediates in the biosynthesis of ACV.

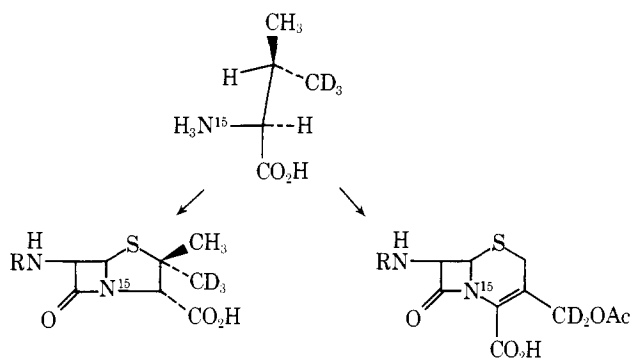
Exposure of mesaconic-*methyl*-*d*<sub>3</sub> acid<sup>7</sup> to  $\beta$ -methylaspartase<sup>8</sup> in the presence of <sup>15</sup>NH<sub>3</sub><sup>9</sup> afforded L-*threo*- $\beta$ -methylaspartic-<sup>15</sup>N,*methyl*-*d*<sub>3</sub> acid in 53% yield, which was transformed into (2*S*,3*S*)-methylvaline-<sup>15</sup>N,3-*methyl*-*d*<sub>3</sub><sup>10</sup> via the sequence of reactions previously described<sup>6</sup> in an overall yield of 23%.

After incubation of (2*S*,3*S*)-methylvaline-<sup>15</sup>N,3-*methyl*-*d*<sub>3</sub> with washed cells of *C. acremonium* mutant C91<sup>11</sup> for 10 hr, the resulting penicillin N and cephalosporin C were isolated<sup>6</sup> and subjected to mass spectrometric analyses<sup>12</sup> as their *N*-benzoylpenicillin N methyl ester and *N*-acetylcephalosporin C methyl ester derivatives,<sup>7</sup> respectively. The most intense mass fragments at *m/e* 174<sup>13</sup> and 230, possessing the valinyl moieties of penicillin N and cephalosporin C, respectively, were selected for the calculation of isotopic ratios. The data in Table I clearly demonstrate that the entire skeleton<sup>14</sup> of (2*S*,3*S*)-methylvaline-<sup>15</sup>N,3-*methyl*-*d*<sub>3</sub> is incorporated intact into penicillin N and cephalosporin C. This conclusion is vindicated by the very prominent (P + 4) peak at *m/e* 178 and the (P + 3) peak at *m/e* 233, respectively. It is evident that the valine molecule underwent

**Table I.** Isotopic Distribution of Cephalosporin C and Penicillin N Derivatives Derived from (2*S*, 3*S*)-Methylvaline-<sup>15</sup>N, 3-methyl-*d*<sub>3</sub>

 <i>m/e</i> 230		 <i>m/e</i> 174	
<i>m/e</i>	Ceph C	<i>m/e</i>	Pen N
230	63.2	174	63.4
231	6.5	175	2.0
232	24.1	176	1.6
233	6.2	177	27.6
		178	5.4

transamination to a considerable extent as indicated by the appearance of (P + 3) and (P + 1) peaks for penicillin N and (P + 2) and (P + 1) peaks for cephalosporin C. As the isotopic ratios of the valinyl moieties in both penicillin N and cephalosporin C were found to be very similar, this finding is consistent with the proposition that both  $\beta$ -lactam antibiotics originate from a common tripeptide intermediate such as ACV.



We next turned our attention to the question whether  $\alpha,\beta$ -dehydroamino acid units are involved during the incorporation of L-valine into ACV, since an  $\alpha,\beta$ -dehydrovaline derivative of a tripeptide has been proposed as a possible common intermediate in the biosynthesis of penicillin and cephalosporin antibiotics.<sup>15-17</sup> As cell extracts of *C. acremonium* incorporated only small quantities of L-valine-<sup>14</sup>C into ACV, uniformly labeled L-valine-<sup>14</sup>C (20  $\mu$ Ci)<sup>18</sup> was exposed to washed starved cells of *C. acremonium*<sup>11</sup> for 15 min to yield ACV-<sup>14</sup>C (0.36  $\mu$ Ci), which was isolated as its sulfonic acid derivative.<sup>2</sup> After acid hydrolysis, the radioactive valine (0.19  $\mu$ Ci) was isolated and its absolute configuration was confirmed to be that of D configuration by use of L- and D-amino acid oxidases.<sup>11</sup> In a similar fashion, L-[2,3-<sup>3</sup>H]valine (40  $\mu$ Ci)<sup>18</sup> was fed and the resulting tritiated ACV (0.4  $\mu$ Ci) was isolated and hydrolyzed with acid. The tritiated valine (0.2  $\mu$ Ci) thus produced was again exposed to L- and D-amino acid oxidases. The results clearly show that only the tritium atom at C-3 of the L-[2,3-<sup>3</sup>H]valine is retained during its incorporation into ACV, for the tritium content of the valinyl residue (7000 cpm), derived from ACV, was about the same as that of the  $\alpha$ -ketovaleric acid (6750 cpm), obtained upon oxidation with D-amino acid oxidase. Although the intimate details of the epimerization of the  $\alpha$ -center of valine remain to be established, our data are incompatible with reaction mechanisms involving  $\alpha,\beta$ -dehydrovalinyl intermediates or direct internal hydride shift, common among hydroxy acid racemases.<sup>19</sup>

These results, and the observation that tritiated ACV, labeled at the  $\alpha$ -center of the D-valinyl moiety, was incorporated into penicillin N by protoplast lysates of *C. acremon-*

*ium* with retention of tritium,<sup>5</sup> provide cogent evidence against the participation of free  $\alpha,\beta$ -dehydrovalinyl tripeptide intermediate(s) in the formation of the penam nucleus.

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## Chemiexcitation Mechanisms. The Role of Symmetry and Spin-Orbit Coupling in Diradicals

Sir:

For a Woodward-Hoffmann symmetry-forbidden pericyclic reaction, Figure 1a shows schematically the three potential energy surfaces connecting the ground (G), lowest